

SHENTEK

Vesivirus 2117 Detection Kit

User Guide

Version: A/1

For Research Use Only

Product No.: 1506743

Reagents for 50 Reactions

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(IMPORTANT: Please read this document carefully before experiment.)

1. Product information

■ Product description

The SHENTEK® Vesivirus 2117 Detection Kit is designed for qualitative detection of Vesivirus 2117 contamination in mammalian cell lines such as CHO cells used in biopharmaceutical production processes. The kit works together with SHENTEK® Virus DNA & RNA Extraction Kit (Product No.1506730).

The detection assay could reach a sensitivity of 50 copies per reaction, and shows high specificity without cross-amplification with bovine, porcine, and engineering cell lines (CHO, VERO, 293T, MDCK, NS0, and Sf9) or engineering bacteria (*E. coli*, *Pichia pastoris*) genomes. Vesivirus 2117 Positive Control and VIR Internal Control (IC) are provided to ensure reliable results. Combined with SHENTEK® Virus DNA & RNA Extraction Kit, these kits can efficiently recover and detect Vesivirus 2117 RNA from complex sample matrix such as cells and serum.

The product described herein is for research use only and not for diagnostic or therapeutic use.

■ Kit components and storage

WARNING: Read the Material Safety Data Sheets (MSDS) and follow the handling instructions. Wear appropriate protective eyewear, clothing, mask and gloves.

Table 1. Kit components and storage

Reagent	Part No.	Quantity	Storage
VIR Internal Control (IC)	NNA057	600 µL × 1 tube	-20°C
5×RT-qPCR Buffer	NNB018	300 µL × 1 tube	
RNase-Free H ₂ O	NND008	1.2 mL × 1 tube	
RT-qPCR Enzyme MIX	NNC079	100 µL × 1 tube	-20°C, protect from light
VV-2117 Primer&Probe MIX	NNC123	170 µL × 1 tube	
VIR IPC MIX	NNC107	100 µL × 1 tube	

Note: According to USP <659>, items with recommended storage temperatures not exceeding -20°C shall be stored within $\pm 10^{\circ}\text{C}$ (-30°C to -10°C). Short-term storage below -30°C is acceptable when supported by stability data.

The kit components can be stored under the appropriate conditions for up to 24 months. Please check the expiration date on the labels.

■ **Applicable instruments, including but not limited to the following**

- SHENTEK-96S Real-time PCR System
- LightCycler 480 II Real-Time PCR System
- ABI 7500 Real-Time PCR System

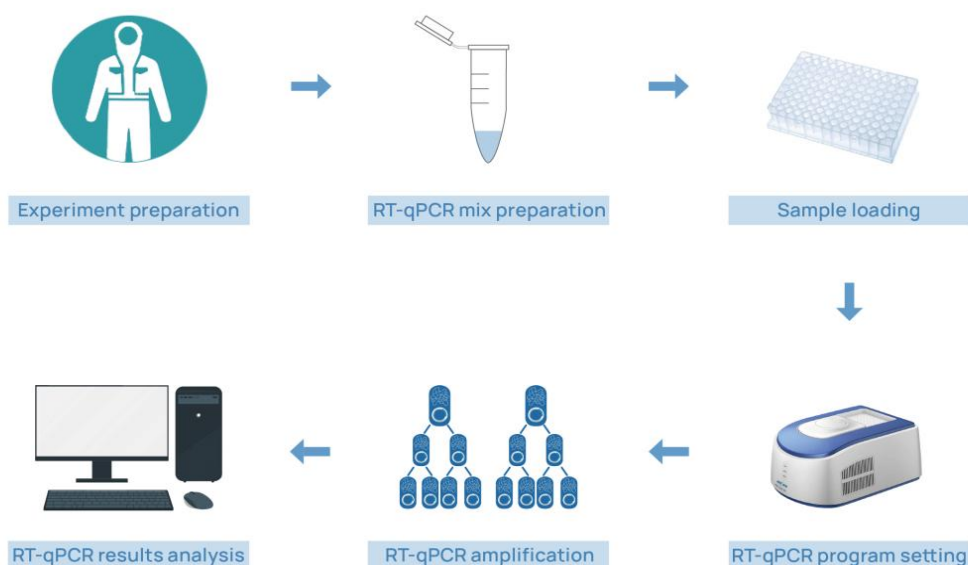
■ **Required materials not included in the kit**

- Low retention, RNase/DNase-free, sterile microcentrifuge tubes: 1.5 mL or 2.0 mL
- 96-well qPCR plates with sealing film or 8-tube PCR strips with caps
- Low retention, sterile filter tips: 1000 μL , 200 μL , 100 μL and 10 μL
- SHENTEK[®] Virus DNA & RNA Extraction Kit (Product No. 1506730)
- SHENTEK[®] Vesivirus 2117 Positive Control (Product No. 1506743-R01)
- Pretreatment buffer (NND002), Proteinase K Buffer (NND025), Proteinase K (NND023), Precipitation solution I (NND003) and 5M NaCl (NND040), components of the Virus DNA & RNA Extraction Kit (Product No. 1506730)

■ **Related equipments**

- Benchtop microcentrifuge
- Microplate and microtube shaker
- Vortex mixer
- Real-Time PCR System
- Pipettes: 1000 μL , 200 μL , 100 μL and 10 μL

■ Workflow



2. Methods

■ Experiment preparation

1. Wear appropriate protective eyewear, mask, clothing and gloves.
2. Please handle the positive control and the extraction procedure in a biosafety cabinet and follow the BSL-2 laboratory safety guidelines.
3. Use consumables and reagents that are certified nuclease-free.
4. Thaw the kit completely at 2-8°C or melt on ice, vortex and spin briefly.

■ Sample pretreatment

Use Virus DNA & RNA Extraction Kit for Vesivirus 2117 extraction. Here is a brief introduction to the process. For detailed steps, please refer to the extraction user guide (Product No. 1506730).

1. Preparation for test samples

For cell culture and bovine serum samples

- (1) Collect 100-200 μL of cell culture ($\leq 10^7$ cells) or bovine serum sample and add to a new 1.5 mL microcentrifuge tube.
- (2) Add Pretreatment buffer (NND002) to the sample at the ratio of 1:10, and vortex to mix well. Incubate at room temperature (RT) for 5 minutes (min).

- (3) Centrifuge at 450×g for 30 min at 4°C, and carefully transfer the supernatant to a new microcentrifuge tube.

2. Preparation of control samples

Positive Control Samples (PCS)

Take one Vesivirus 2117 Positive Control and follow the same extraction procedure used for the test samples.

Negative Control Samples (NCS)

Take an equal volume of RNase-Free H₂O or 1×PBS as the test sample and follow the same extraction procedure used for the test samples.

3. Samples digestion

- (1) Add 100 µL Proteinase K Buffer (NND025) and 50 µL Proteinase K (NND023) to each sample.
- (2) Add 10 µL VIR Internal Control (IC), vortex and centrifuge briefly for 3-5 seconds (s).
- (3) Incubate the samples at 55°C for 30 min.
- (4) Add 10 µL 5M NaCl (NND040) and 10 µL of diluted Precipitation solution I (NND003). (Please check the following user guide before each use of the kit: Precipitation solution I dilution of Virus DNA & RNA Extraction Kit User Guide).
- (5) Vortex and centrifuge briefly for 3-5 seconds (s).

4. Extraction of viral nucleic acid

Automated extraction is recommended using rHCDpurify system for viral nucleic acid extraction, and follow the user guide of SHENTEK® Virus DNA & RNA Extraction Kit.

■ RT-qPCR MIX preparation

1. Determine the number of reaction wells according to the number of test samples and control samples with 2 replicates for each sample generally.

Number of reaction wells = (1 No Template Control (NTC) + 1 Negative Control

Sample (NCS) + 1 Positive Control Sample (PCS) + test samples) × 2

2. Prepare RT-qPCR MIX according to the following table.

Table 2. RT-qPCR MIX preparation

Reagents	Volume/reaction		Volume for 36 reactions (includes 10% overage)
	Test samples	NTC	e.g. Test samples
5×RT-qPCR Buffer	4 µL	4 µL	160 µL
RT-qPCR Enzyme MIX	1 µL	1 µL	40 µL
VV-2117 Primer&Probe MIX	3.3 µL	3.3 µL	132 µL
VIR IPC MIX	1.7 µL	1.7 µL	68 µL
VIR Internal Control (IC) *	/	0.5 µL	/
Total volume	10 µL	10.5 µL	400 µL

Note: If VIR Internal Control (IC) has not been added during sample extraction, 0.5 µL VIR Internal Control (IC) should be added to the RT-qPCR MIX.

■ RT-qPCR Reaction MIX preparation

1. Vortex each solution to mix well. Then add the solution to each well according to Table 3, and refer to 96-well plates layouts shown in Table 4.

Table 3. RT-qPCR Reaction MIX preparation in each well

NTC (No Template Control)	10 µL RT-qPCR MIX + 10 µL RNase-Free H ₂ O
NCS (Negative Control Sample)	10 µL RT-qPCR MIX + 10 µL of purified NCS
PCS (Positive Control Sample)	10 µL RT-qPCR MIX + 10 µL of purified PCS
Test Sample	10 µL RT-qPCR MIX + 10 µL of purified test sample

Table 4. Example of 96-well plate layouts

NCS	NCS				S1	S1				PCS	PCS	A
					S2	S2						B
					S3	S3						C
					S4	S4						D
					S5	S5						E
												F
												G
NTC	NTC											H
1	2	3	4	5	6	7	8	9	10	11	12	

Note: This example shows 1 NTC, 1 NCS, 1 PCS, and five test samples, with duplicate wells for each sample.

Note: In specific testing, the plate layout can be adjusted based on the sample quantity.

2. Seal the 8-tube PCR strips with caps or the 96-well plates with sealing film. Mix the contents of the wells in microplate shaker, then spin down the plate for 10 seconds in microcentrifuge and place it in the qPCR instrument.

■ RT-qPCR program setting

Take the SHENTEK-96S Real-time PCR System (software version 8.2.2) as an example, and set the RT-qPCR program as follows:

1. Click on the **Experiment Wizard**.
2. Select Step 1 on the **Well Plate Edit** page: Select the reaction wells.
3. Select Step 2: Select the **VV-2117 Detection** program in the project.
4. Click **Start** to run the program on the **Experimental Run** page.

If you have alternative qPCR system, set up the instruments as follows:

1. Run a new method program and select the quantitative PCR assay template.
2. Run a new Probe template, and type the name **VV-2117**. Select **FAM** in the Reporter Dye drop-down list and select **none** in the Quencher Dye drop-down

list, then click **OK**. Select **VIC** in the Reporter Dye drop-down list and select **none** in the Quencher Dye drop-down list, then click **OK**. Select the detection reference fluorescence as ROX (optional).

3. Set thermal-cycling conditions:
 - a. Set the cycling reaction volume to 20 μ L.
 - b. Set the temperature and the time as follows:

Table 5. RT-qPCR running temperature and time

Step	Temp.	Time(mm:sec)	Cycles
Reverse transcription	50°C	15:00	1
Activation	95°C	00:30	1
Denaturation	95°C	00:15	40
Annealing/extension	60°C*	01:00	

*Instrument will read the fluorescence signal during this step.


■ Result analysis

Take the SHENTEK-96S Real-time PCR System (software version 8.2.2) as an example.

1. Select **step 3** in the **Edit** page: define the reaction wells, set the sample type for the NTC well as no template control, the PCS well as the positive control, the NCS well as the negative control, and the test sample well as unknown.
2. Click **Analysis** on the **Experimental Analysis** page, and the detection values of NTC, NCS, PCS and test samples can be presented in the Reaction well Information Table.

For ABI 7500 Real-Time PCR System instrument (SDS v1.4), please follow the steps below:

1. Select the **Results** tab, then click **Amplification Plot**. In the Analysis Settings window, enter the following settings:
 - a. Select **Manual Ct**.
 - b. In the Threshold field, enter value 0.02.

- c. Click **Analyze**, and check whether it is a specific amplification curve.
2. Select the **Results** tab, and click **Plate** tab, then set tasks for each sample type by clicking on the Task Column drop-down list and then click :
 - a. NTC wells: target DNA detector task = NTC.
 - b. NCS, PCS, unknown samples wells: target DNA detector task = Unknown.
3. Select **File >> Export >> Results**. In the **Save as type** drop-down list, select **Results Export Files**, then click **Save**.

Note: If you use a different instrument or software, refer to the applicable instrument or software documentation. Normally, real-time PCR instrument software will export the data reports automatically.

■ Acceptance Criteria

1. Guidance for the control samples as in the following table.

Table 6. Control sample result analysis

Quality control samples	FAM	VIC
NTC	Undetected or absence of specific amplification for duplicate runs	Ct<35.00 and specific amplification for duplicate runs
NCS	Undetected or absence of specific amplification for duplicate runs	Ct<35.00 and specific amplification for duplicate runs
PCS	Ct<35.00 and specific amplification for duplicate runs	Ct<35.00 and specific amplification for duplicate runs

The criteria for control sample shall be based on method validation data, and considered to satisfy the requirement of DL (Detection Limit).

2. Guidance for test samples as in the following table.

Table 7. Test sample result analysis

FAM	VIC	Conclusion
Ct<40.00 (at least one well) and specific amplification	Ct<35.00 and specific amplification for duplicate runs	Positive
Undetected or absence of specific amplification for duplicate runs	$Ct_{\text{Sample}} - Ct_{\text{NCS}} < 2$	Negative
	$Ct_{\text{Sample}} - Ct_{\text{NCS}} \geq 2$	Not conclusive, and presence of PCR inhibitors*

* Compare the VIC Ct value of the sample to the NCS Ct value. If the difference is greater than 2, it indicates inhibition in extraction or detection. Retest or remove the potential inhibitors appropriately to confirm the virus contamination.

Note: In an event of specific sample or some abnormalities occur, or results are difficult to determine, please contact us for technical support.

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Support & Contact



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